

Home Search Collections Journals About Contact us My IOPscience

Light-based therapy on wound healing: a review

This content has been downloaded from IOPscience. Please scroll down to see the full text. 2014 Laser Phys. 24 083001 (http://iopscience.iop.org/1555-6611/24/8/083001)

View the table of contents for this issue, or go to the journal homepage for more

Download details: This content was downloaded by: piksuan88 IP Address: 161.139.220.12 This content was downloaded on 25/06/2014 at 07:36

Please note that terms and conditions apply.

# **Topical Review**

# Light-based therapy on wound healing: a review

## Lau Pik Suan<sup>1</sup>, Noriah Bidin<sup>1,3</sup>, Chong Jia Cherng<sup>1</sup> and Asmah Hamid<sup>2</sup>

<sup>1</sup> Advance Photonic Science Institute, Faculty of Science, Universiti Teknologi Malaysia (UTM), Skudai 81310 Johor, Malaysia

<sup>2</sup> Department of Biomedical Science, Faculty of Health Science, Universiti Kebangasaan Malaysia, Kuala Lumpur, Malaysia

E-mail: noriah@utm.my

Received 15 July 2013, revised 4 December 2013 Accepted for publication 24 April 2014 Published 18 June 2014

## Abstract

Wound healing is a complex matrix and overlapping process. In order to accelerate the healing process and minimize bacterial infection, light-based therapy was applied to stimulate bio-reaction to improve healing. The aim of this paper is to review the effects induced by light source (laser and incoherent light like LED) on different biological targets. The light-based therapy techniques were categorized according to the wavelength, energy density, type of irradiance and activity of tissues in the healing process. Out of 80 cases, 77% were animal studies, 5% were human studies and 18% were cell studies. Around 75% of light-based therapy has an advantage on tissue interaction and 25% has no effect or inhibition on the healing process. The appropriate dose appears to be between 1 and 5 J cm<sup>-2</sup>. At shorter wavelength, photobiostimulation would be effective with a high frequently administrated low-energy dose. On the other hand, for longer wavelength it is the reverse, i.e., more effective with a low frequent treated schedule and a high-energy dose.

Keywords: wound healing, low-level laser therapy, biophotonic, photobiostimulatory

(Some figures may appear in colour only in the online journal)

## 1. Background

Wounds are divided into two types: internal and external. An internal wound is created due to circulation, neuropathy or medical illness. Skin is a protective barrier for isolation of the body and environment. Breaking the barrier allows the creation of an external wound such as an incision, trauma or burn [1]. The external wound can repair itself via the normal healing process. This involves four stages including hemostasis, inflammation, proliferation and remodeling [2–4]. The healing time depends on the size of the wound, but the healing rate is an independent process [5]. When the barrier protection is open and exposed to the environment, bacteria are allowed to build up and breeding occurs in the wound site. Clinical

treatment is needed in this case. In conventional treatment, medicine is applied to the wound such as antibiotics, dressing and ointment to prevent bacterial infection, activate metabolism and reduce wound pain [6]. Nowadays, laser therapy is attracting research applying it to *in vitro* and *in vivo* targets. Laser therapy is a drug-free, extremely safe, easy to apply procedure and complements many traditional therapies. Hence, it is much better than conventional treatment. Most importantly, laser therapy can minimize the risk of bacterial infection and thus accelerate the healing process.

## 2. Low-level laser therapy (LLLT)

Laser therapy was first demonstrated by Endre Mester in 1967. The experiment in hair growth of mice revealed unexpected

<sup>&</sup>lt;sup>3</sup> Author to whom correspondence should be addressed.

results that led towards a new discovery in using laser treatment [7]. Laser therapy involves the application of a low-power laser that is irradiated on a wound in order to stimulate the healing process. It is also referred to as low-level laser therapy (LLLT) [8, 9]. The important mechanism in laser therapy is photobiostimulation. LLLT is a biophotonic technique which stimulates the biological cell through the absorption of photons [10–12]. Photobiostimulation research has been reported since the introduction of lasers in the biological field and clinical applications [13–17]. Nevertheless, the exact mechanism of LLLT is not yet fully understood. Theoretically, the laser is absorbed by a light agent such as the mitochondria, hemoglobin and melanin. Once the tissue has absorbed the light, an electron in lower orbit will be excited and produce internal conversion energy. Inter-cellular communication is motivated by bio-reaction and restores the normal cell function [17]. Laser therapy reduces the inflammatory reactions, increases collagen deposition and induces greater proliferation [18, 19].

The effects of lasers will depend on parameters like power density and exposure time. The possible result might be to stimulate or inhibit the healing process [20]. Several studies have been carried out utilizing different types of lasers. The absorption relies on light properties such as monochromatism, non-coherence and polarization as well as depending also on the energy gap of the bio-molecule [21, 22]. The intensity of light also affects the outcome of the laser treatment. High power lasers are commonly used to cut through tissue. They are suitable in surgery for cutting or cauterizing. Exposure to a highpower laser for treatment has limitations, as over-exposure will cause disaster. Low-power lasers will stimulate tissue repair through a process of photobiostimulation. These low-level lasers do not have enough power to damage tissue, consequently a heating effect, damage to the skin and side effects may not occur [23]. Different parameters used in various studies raise a lot of complications and difficulties for comparison. Laser therapy has many biological effects either on in vivo or in vitro samples. Amongst them are accelerated tissue repair and cell growth, reduced fibrous tissue formation, anti-inflammation, analgesia, improved vascular activity and increased metabolic activity. The LLLT treatment has been accepted among health care practitioners. However, there is still a lack of documentation regarding the application of LLLT.

The aim of this work is to review the effects of LLLT and its consequences. The dependence on the energy density, types of irradiance and activities of tissue in the healing process after irradiance with LLLT will be discussed in detail. In general, typical medical lasers operate in the wavelength range between ultraviolet and infrared. The spectrum of ultraviolet radiation is in the range 180–400 nm. An excimer laser is an example of an ultraviolet laser. Excimer lasers normally have large beam spot size. This is a disadvantage in medical applications due to low power density. Furthermore, due to the shallow optical penetration depth, the excimer laser is normally used in photoablation to remove superficial surface tissue. This is the mechanism for eye surgery to repair astigmatism and myopia, keratomileusis, diabetic retinopathy and microbial keratitis [24–28]. On the other hand, for a long wavelength radiation, it is able to penetrate deeper into the tissue. Thus, the choice of a specific laser wavelength will depend on its penetration depth into the tissue. As the wavelength is increased further into the infrared region, light is absorbed more by water which limits its penetration into the desired tissue [29]. LLLT can be operated in continuous or pulse mode.

#### 3. LLLT targets and laser sources

Various biological targets and laser sources have been reported for wound healing treatment. 77% of wound healing studies used animals as targets, 4.8% were performed on humans and 18% on cells. 87.5% of the animal studies used rats as experimental subjects. The wavelength of laser used in the treatment covered the range from visible (470 nm) to infrared. Nearly 91% of the irradiance is in the form of continuous wave (cw) and the other 9% in pulse mode. Some reports claimed that the laser irradiation might enhance, inhibit or have no effect on *in vivo* or *in vitro* targets. Around 75% of laser therapies have a positive effect on tissue interaction and 25% have no effect or inhibit the healing process.

The most popular wavelength in LLLT is 632.8 nm, followed by 670 nm (as shown in figure 1). Other lasers, including diode laser at 904 nm, CO<sub>2</sub> (10.6 $\mu$ m), Nd:YAG (1.064 $\mu$ m), and ND:YLF (1.047 $\mu$ m), are commonly employed as a source of illumination. In other cases, a combination of two or more different wavelengths is also used for wound treatment. Almost 60% of the combination techniques comprised short wavelength (632.8 nm), and long wavelength (904 nm).

Energy (J) or energy density (J  $\text{cm}^{-2}$ ) is often used as an important parameter to describe the LLLT performance. Energy density is expressed as follows [22]:

Energy density 
$$(J \text{ cm}^{-2}) = \frac{\text{Output power}(W) \times \text{time}(s)}{\text{Beam area}(\text{cm}^2)}$$
. (1)

The range of energy density normally applied for treatment varies from 0.1 to 140 J cm<sup>-2</sup>. LLLT often operates within 1-5 J cm<sup>-2</sup> and it occupies almost 60% in the case studied. The frequent doses are 1-4 J cm<sup>-2</sup>. The high-energy dose is considered in the range of 15-20 J cm<sup>-2</sup>.

Wound parameters are an important aspect to be considered in order to measure the progress of wound healing. This is the way to quantify and monitor the healing progression. There are several familiar variables to estimate the rate of healing, such as wound contraction size, histology, tensile strength, blood flow and scoring system. In this study, 58% of researchers used histology to establish a wound healing progress. A scoring system for the histological assessment of wound healing is commonly performed by edema, leucocytes, macrophages, granulation tissue, fibroblasts, collagen and epithelialization [30–39].

#### 4. Types of light source

#### 4.1. Visible violet-blue-green-yellow laser

The case studies in this spectral range are listed in table 1. The wavelengths covered are within the range 442–532 nm.



Figure 1. Percentage of tested wavelength for photobiostimulation in wound healing.

Authors	Wavelength (nm)	Target	Energy or power density (J cm <sup>-2</sup> )	Outcome				
Guffey et al [46]	405 470	S. aureus P. aeruginosa	1 3 5 10 15	405 nm effectively kill <i>S. aureus</i> and <i>P. aerugin</i> 470 nm light effectively killed <i>P. aruginosa</i> at all dose levels, but only killed <i>S. aureus</i> at 10 and $15 \text{ J cm}^{-2}$				
Al-Watban <i>et al</i> [40]	442 488+514.5 632.8 780 830	SD rat	20 (all) 19 (Ar)	All lasers were better than control but He–Ne was most effective and He–Cd was least effective, three times weekly				
Adamskaya <i>et al</i> [43]	470 (LED) 630 (LED)	Rat	30	470 nm light significantly influences wound healing				
Al-Watban <i>et al</i> [42]	(488–514) Ar; (670) krypton	SD rat	20 80 100 140	20 J cm <sup>-2</sup> Ar laser (488 nm) was most effective in wound healing; $140 \text{ J cm}^{-2}$ Kr was inhibitive				
Poon <i>et al</i> [44]	532 Q-switch; Nd:YAG	Human fibroblast	0.8	Significant delay in collagen remodeling activity and increase in SCF and b-FGF content				

Table 1	. Wound	treatment	with	visible	violet-	-blue-	-green-	-yellow	laser
---------	---------	-----------	------	---------	---------	--------	---------	---------	-------

Al-Watban et al [40, 41] have compared the wound treatment by using various light sources including 442, 488, 632.8, 780 and 830 nm by fixing the energy density at 20 J cm<sup>-2</sup> and treatment schedule. Most of the results have revealed stimulation in wound healing as compared to the control. He-Ne with dose of 20 J cm<sup>-2</sup> has been attributed the most effective treatment, whereas the He-Cd violet-blue laser was the least. This meant the red laser was better for treatment than the violet-blue laser. The Al-Watban group [42] has conducted a similar investigation by using argon blue-green laser (488-514 nm) and krypton red laser (670 nm). They have found that the argon laser with dose 20J cm<sup>-2</sup> has shown more effective treatment than the krypton laser. Furthermore, zero biostimulation at dose of 80-100 J cm<sup>-2</sup> was realized but inhibition occurs at 140 J cm<sup>-2</sup>. Consequently, the bluegreen laser is more appropriate for wound treatment than the

3

red laser. This is contradictory with previous results [40]. Adamskaya *et al* [43] have conducted wound treatment by combining two light emitting diodes (LED). The LED was comprised of blue light (470 nm) and red light (630 nm) that was used to treat an excision rat model. Significantly, blue light contracted the wound size better than and enhanced epithelization compared to red light.

Poon *et al* [44] have accomplished *in vitro* biostimulation of dermal fibroblasts by using 532 nm Q-switched Nd:YAG. The laser was operated at a maximum energy of 200 mJ per pulse with pulse duration of 4 ns, in various modes including single shot and repetitive at the rates of 1, 2, 5 and 10 Hz. There is no significant difference in collagen synthesis. But the delay in collagen remodeling activity between the stimulated fibroblasts and controls is noticeable. Therefore, wound healing may be delayed without typical clinical features of infection [45]. *Staphylococcus aureus* and *Pseudomonas aeruginosa* are the common aerobes found in the human tract and on the skin. They commonly cause skin infections and damage to tissues. A blue laser is usually used to kill bacteria *in vitro* such as *Staphylococcus aureus* and *Pseudomonas aeruginosa*. LED light at lines 405 and 470 nm is effective on bacteria such as *Staphylococcus* and *Pseudomonas aeruginosa* [46]. The LED of 405 nm is capable of destroying *Pseudomonas aeruginosa* and *Staphylococcus aureus* achieving a killing rate of 95 and 90%, respectively. All dose levels of 470 nm light are capable of destroying *Pseudomonas aeruginosa*, whereas bacteria *Staphylococcus aureus* can only be destroyed at 10 and 15 J cm<sup>-2</sup>. Thus, blue light can produce a bactericidal effect.

#### 4.2. Visible red laser

In this section, 40 cases have been studied. The range of lasers used in this wound treatment are listed in table 2. Almost half of them exploit lasers at 632.8 nm; this is the most popular wavelength used to expose wounds. Kana et al [47] have applied low and high energy density lasers for the wound treatment. They used energy densities of 4 and 20 J cm<sup>-2</sup> and daily exposure on rat wounds with 632.8 nm. The low irradiation energy density showed a faster contracted wound area than the control group. This indicates that the energy density of 4J cm<sup>-2</sup> is capable of accelerating wound healing. In contrast, the high dose irradiation has no effect on the healing process. This finding is matched by Mester's result [7]. Other related work is also reported by Surinchak et al [48] and Lyons et al [49]. Both groups have claimed that LLLT is capable of breaking the strength of rat skin following irradiation with energy not exceeding  $4.5 \,\mathrm{J}\,\mathrm{cm}^{-2}$ .

The types of samples also influence the outcome of wound treatment. No significant effects are realized when rabbit, swine and horse were used as samples [48, 50-52]. Ghamsari et al [53] have studied the effect of He–Ne on suture wounds of the teat in dairy cattle. The collagen fibers and tensile strength changed quite significantly after being treated by He-Ne radiation. Different species of animals may have different skin components, hence the characteristics of penetration depth of cellular molecules also differ. Some studies indicate that linearly polarized light can survive to propagate through long distances in biological tissue [54]. Al-Watban et al [41] utilized linear polarized He-Ne to cure an artificial wound with energy density in the range up to  $60 \text{ J} \text{ cm}^{-2}$ . The acceleration of 27% in healing time and 49% in contraction area of wound was achieved after administering a dose of 25 J cm<sup>-2</sup>. Al-Watban et al [40-42, 55-60] claimed that acceleration in wound healing is dependent upon the dose. The rate of healing process will increase up to a certain stage. The stimulatory effect was observed to decrease and inhibit beyond the optimum level. Photobiostimulation effect was dependent on the energy density of either in vivo or in vitro target. In vitro cellular studies [61–64] using human skin fibroblasts, and He–Ne lasers at various energy densities were utilized. The results were sufficient to produce measurable changes causing an increase in procollagen production. Although these findings indicate that the human skin fibroblast had displayed optimum effect at dose  $5 \text{ J cm}^{-2}$ , no stimulatory effect is revealed at a dose lower than 0.5 J cm<sup>-2</sup>. Meanwhile, cellular damage occurs after the given dose is exceeded by 10 J cm<sup>-2</sup>. High doses are usually used in photodynamic therapy [10].

Photobiostimulation is not only dose-dependent, but also wavelength-dependent. In the 20th century, most research was conducted using wavelengths 635, 636 and 660 nm. The results showed a lack of stimulatory action either in low or high dosages [65–68]. The 670 nm laser was developed in the 20th century. The stimulatory function is understood to be different compared to a wavelength in the range 635–660 nm. An opposite finding is noted, whereby the healing process becomes more effective at a lower dosage.

Puglzese *et al* [69] have exploited GaAlAs laser at 670 nm with an output power of 9 mW. A Wistar rat was used as target. Laser radiation with low energy showed a stimulating effect on the target. An inhibiting effect is obtained after exposure to high-energy radiation. Referring to table 2, 4J cm<sup>-2</sup> is the dominant dosage to enhance the healing process, with the results in good agreement with several other researchers [69–71].

#### 4.3. Infrared laser

Infrared radiation is divided into three categories: nearinfrared ( $0.8-1.5\mu$ m), middle-infrared ( $1.5-5.6\mu$ m) and far-infrared radiation ( $5.6-10000\mu$ m). Near, middle and far-infrared rays have different photobiological effects [72]. Near-infrared wavelengths are weakly absorbed and penetrate deeply into the tissue (this penetration is, however, limited by optical scattering). In the middle- and far-infrared, water absorbs intensely, with light then only having very superficial effects [73]. The schematic diagram in figure 2 illustrates the effect of an infrared beam on tissue.

As the laser penetrates and is absorbed by soft tissue, a stimulatory effect occurs due to the activation of adenosine triphosphate (ATP) through bio-reaction. Table 3 summarizes the studies that involve infrared laser to stimulate the target. Grossman *et al* [74] conducted treatment by using a near-infrared diode laser at 780 nm with an output power of 6.5 mW. Proliferation of a culture of normal human keratinocytes *in vitro* was studied at various energy densities from 0 to  $3.6 \text{ J cm}^{-2}$ . Proliferation *in vivo* condition was also conducted in order to accelerate wound healing. The 780 nm-irradiation was claimed to induce a positive effect on wound healing.

LLLT has shown a variety of effects including increased maturation of collagen, fibroblasts and capillary vessels. The treatment was also capable of reducing pain and decreasing inflammation [75–79]. Laser therapy has been carried out using near-infrared at 830 nm with an output power of 10–40 mW. The healing process has shown a stimulation effect when treated with low energy density not exceeding 5J cm<sup>-2</sup> but tends to inhibit as the doses approach 20J cm<sup>-2</sup> [77, 80–82].

The healing process was also studied by comparing LLLT treatment with other method. As an example, comparisons between laser and ultrasound have been reviewed. A GaAs laser at 830nm with an output power of 30mW was utilized. The sample was exposed daily to a laser for interations of

## **Table 2.** Wound healing studies involving red laser wavelength.

Authors	Target	$\lambda$ (nm)	Energy density, (J cm <sup>-2</sup> )	Power (mW) (*mW cm <sup>-2</sup> )	Schedule	Outcome
Kana [47]	Rat	632.8	4		Daily	Increased collagen synthesis
Surinchak [48]	Rabbit	632.8	20 1.1	_	Every third day	Inhibit No significant effect
Surinchak [48]	Rat	632.8	2.2 2.2 4.5	_	Twice daily	Increased 55% breaking strength
Hunter [50]	62 swine	632.8		64*	—	No significant effect
Lyons [49] Frets [51]	Rat 8 horse	632.8 632.8	1.22 45.9	1.56 13	Every other days	Enhanced healing No significant effects
Al-Watban [41]	SD rat	632.8	20	10.5*	6 times weekly	Accelerated healing
			25 60			Optimum dose in healing Inhibit healing
Atabey [52]	38 rabbits	632.8	3.8	5	15 min daily	No significant wound contraction
						but epidermal thickening, increased
Al-Watban [40] Ghamsari [53] Nunes [100] Hawkin [10]	SD rat 16 dairy cattle 15 rat Human skin fibroblast	632.8 632.8 632.8 632.8	20 3.64 1 0.5 2.5	8.5 10	3 times weekly 30s for 10d Daily for 3d Single dose	fibroblast and dermal vascularity Enhanced healing Enhanced healing No significant effect Increased fibroblast Increased fibroblast
Hawkin [61]	Human skin fibroblast	632.8	5 10 0.5 2.5	3*	2 d	Increased fibroblast Cellular damage No significant effect Increase in chemotaxis–chemokinesis
Rabelo [101]	50 rat	632.8	5 10 16 10	15	17 s	and haptotaxis Optimum dose, enhanced healing Worse, DNA damage Worse, DNA damage Enhanced healing, less intense
Vasukawa [102]	SD rat	632.8		85	Every other day	inflammatory Better than control
		(22.0	Ē	17.0		Optimum dose, enhanced healing
Houreid [62]	fibroblast	032.8	5		Day 1 and day 4	damage and no cytotoxicity
Evans [63]	Human skin	632.8	16 5 16	18.8	Daily	Inhibit and damage Enhanced healing No significant effect
Houreld [64]	Human skin	632.8	5	_	_	Improved wound healing
Hedge [103]	fibroblast 105 rat	632.8	1 2 3	7	Single irradiance	Lowest effect Low effect Optimum dose Similar with optimum effect
			5			Low effect in laser group
Nussbanm [65]	70 SD rat	635	$\frac{1}{20}$		3 times weekly	No significant effects
Sekhejane [66]	Human skin	636	5	95	Single dose	Enhanced healing
Walker [67]	36 mice	660	0.5 1.5	15	3 times weekly	No significant effect No significant effect
Gonzaga [ <mark>68</mark> ]	24 rat	660	4 20		7 d	No significant effect Facilitates myofibroblast and
Al-Wathan [42]	SD rat	670	140	_	_	proliferation Worse
Medrado [70]	72 Wistar rat	670	4	9	Single dose	Enhanced healing
Puglzese [69]	72 Wistar rat	670	8 4	9	Single dose	No significant effect Enhanced healing
Do Nascimento	18 Wistar rat	670	8	2	7 d	No significant effect Enhanced healing
[104]				15		Enhanced healing
C-1 [105]	40	(70	20	25	Delle	Optimum dose
Oai [103] Medrado [106]	49 rat 112 Wistar rat	670	1	9	Daily	Enhanced healing

(Continued)

Table 2. (Continued)								
Authors	Target	$\lambda$ (nm)	Energy density, (J cm <sup>-2</sup> )	Power (mW) (*mW cm <sup>-2</sup> )	Schedule	Outcome		
Reis [71]	32 rat	670	4	9		Enhanced healing		
de Oliveira	50 Wistar rat	670	4	—	—	No significant effect		
Guirro [107]			7			No significant effect		
Pinheiro [93]	30 Wistar rat	685	20	_	Every other day for 7 d	Enhanced healing		
			40			Worse		
Rodrigo [80]	36 Wistar rat	685	20	30	Single dose	Worse		



Figure 2. Laser tissue interaction—various wavelengths will reach different depths into tissues.

1 min or an energy density of  $0.5 \text{ J} \text{ cm}^{-2}$ . Meanwhile, treatment with ultrasound (2 ms on; 8 ms off) was performed by exposing the sample using a power density of  $0.1 \text{ W} \text{ cm}^{-2}$  for a duration of 5 min daily. Laser and ultrasound treatment have shown significant effects as compared to control group. Nevertheless, the ultrasound group did not achieve a statistically significant effect on wound healing [77].

GaAs laser at 904 nm is a popular light source in the infrared region. A continuous 904 nm GaAs was employed as a source for treatment. The energy density of 1J cm<sup>-2</sup> was exposed for 10 min per day. The treatment was carried out for 10 d. The finding was an increase in fibroblasts and wound breaking, i.e. it was beneficial for proliferation, as decreasing the macrophage and PNL means decreasing the inflammatory duration [79].

GaAlAs diode laser at 980 nm is also used for stimulating wound healing processes. In this study, the laser was operated at variable power within the range 1.5–10W. With such a long wavelength and high power laser, the beneficial effect can only be achieved after a short interaction time per treatment for long interval times [83, 84]. Table 4 lists the variation of powers and days of treatment using 980 nm. The interaction time for every treatment is kept constant at 1 s.

Histology was analysed including epitheliazation, cellular content, granulation tissue, collagen deposition and vascularity by microscopic observation. Treatment with 5W for 2d was found to significantly enhance the healing process [84]. LLLT will regenerate the lymphatic system during the process of healing, which affects the occurrence of oedema and adhesion. The quicker the

fluid waste products oedema can be drained, the better the potential of wound healing. The lymphatic system is primarily responsible for the evacuation of this oedema. If the lymphatic system is destroyed by the incision, then the regeneration process of these lymph vessels will determine the evolution of the scar [85].

A number of studies have reported that longer wavelengths (in the far-infrared region) are also able to motivate the bioreaction to accelerate the healing process [86–88]. Nd:YAG laser in pulse mode, for example, has been employed in the wound healing process. Two pulse modes have been conducted: 20 pulses per second (pps) with power of 1.75 W and 30 pps with power of 3 W, respectively. Differences in the distribution of matrix proteins during healing and the coagulation of the tissues were revealed after exposure to low energy laser treatment. This explained the minimal scarring, contraction and pigmentation of the lasered tissues as compared to conventional incisions [89]. The success of Nd:YAG laser pulse for stimulating the matrix process during healing does not mean that this will also apply for the continuous mode.

Ribeiro et al [86] used human skin fibroblast cells and irradiated with variable lasers including 632.8, 830 and 1064 nm at low energy density level. This study showed that cells had a high degree of haptotaxis and migration as well as ATP luminescence at 632.8 nm but no response at 1064 nm. This in contrast with previous claims that far-infrared is also capable of accelerating the healing process. In order to ensure this claim, another longer wavelength laser was investigated. In this case, de Freitas et al [90] used CO<sub>2</sub> gas laser to quantify statistically the myofibroblasts and compared with conventional treatments in the rat model. The result confirmed that the conventional treatment showed an increase in the number of myofibroblasts during the healing which is far better than  $CO_2$  laser therapy. The threshold energy density and intensity are biologically independent parameters. The independence is of practical importance, at least for medical applications. Clearly, photobiological effects are more dominant at a low energy density level based on the account of the success and the failure in most of the cold laser uses since Mester's pioneering work [91, 92].

#### 4.4. Alternatives to laser light source

Research in this domain mostly covers low-level laser studies; however, due to the high cost and safety aspect there is a need to consider other alternative light sources [76]. Recently, LED

Topical Review

<b>Table 3.</b> Wound-healing studies involve infrared laser.									
Author	Target	$\lambda$ (nm)	Energy density	Power (mW) (*mW cm <sup>-2</sup> )	Schedule	Outcome			
Al-Watban [40] Grossman [74]	SD rat Culture normal	780 780	20 0–3.6	6.5	3 times weekly Single dose	Improved healing Enhanced proliferation			
Arcangelo [108]	human keratinocytes 24 Wistar rat (hard palate)	808	_	4000 6000		Worse Worse			
Gungormus [109] Souil [110]	18 Wistar rat Rat	808 815	10	4000 1500 (pulse)		Improved healing Improved healing			
Petersen [111] Vinck [76]	6 horse Fibroblast (old chicken embryos)	830 830	2 1	40	Daily 3 d	No effects Improved healing (*no different with LED			
Mendez [75]	60 Wistar rat	830		_	_	Increased maturation collagen but no reduced			
Lanzafame [82] Rezende [81]	Rat 48 rat	830 830	5 1.3	60	 Single dose	Improved healing Optimum dose for			
Rodrigo [80] Tikiz [77] Lowe [112]	36 Wistar rat 32 Wistar rat 50 mice	830 830 890	3.0 20 0.5 0.18 0.54 1.45	50 30	Single dose 3 times weekly Pulse 270 Hz	Improved healing No effects Worse Improved healing No effects No effects Optimum dose for			
Ezzati [113]	67 rat (burn)	890	2.3		Pulse 3000 Hz	improved healing No effects Improved healing			
Longo [114]	16 rat	904	3	3000 Hz 1500 Hz	5 d	Improved healing No effects			
Skinner [115]	Fibroblast procollagen production	904	0.01–0.5		1-4 d	Optimum dose for healing			
Pereira [116]	NIH 3T3	904	1 (6h interval) 2 (6h interval)		6d	Optimum improved healing No effects			
Demir [79] Herascu [117]	124 mice Patient	904 904	$\frac{1}{2}$ (6 h interval)	$\frac{6}{15}$ 20	$\frac{10d}{D}$	No effects Improved healing Improved healing			
Silveiro [118] Silveiro [119]	Rat 30 Wistar rat	904 904	3 5	15–30 15–30	After trauma 2, 12, 24, 48, 72, 96, 120h	Improved healing Improved healing			
Sanati [120]	30 rat	904 632.8	2 2	*20.6 *31.7	Every other day	Improved healing No effects			
Kawalec [84]	72 mice 72non diabetic mice	980 980	18 36	5000	Every other day Every 4d Every other day	No effects Optimum dose No effects			
Skopin [83]	Fetal human skin	980	_	1500–7500	Every 4d Single dose (2min)	No effects Improved healing			
Skopin [83]	fibroblast Fetal human skin fibroblast	980	_	*73 4500 *73	Single dose 50 s Single dose 2 min	Improved healing Improved healing			
Skopin [83]	Fetal human skin fibroblast	980	_	1500–7500 *97 *120	Single dose (2 min)	No effects Improved healing No effects			
Romanos [89]	Rat	1064		*120 1750 3000	Single dose	Improved healing			
Ribeiro [86]	Rat	1047	1		_	Improved healing with			
Houreld [64]	Human skin fibroblast	632 830	5			632.8 nm beneficial and 1064 nm worse in healing			
Yu [88]	60SD rat	1064 1000–			Single dose, 30 min,	Optimum duration			
Lanbach [87]	12 volunteers inner forearm	12 000 1500			45 min, 60 min Single irradiation	45 min for healing Epidermis recovers fast			

**Topical Review** 

Table 4. Treatment with GaAlAs 980 nm for treatment time of 1 s.

Days of treatment	Energy density (J cm <sup>-2</sup> )
2 4 2 4	18 18 36 36
	Days of treatment 2 4 2 4 4 4

has been favoured as an alternative source for light-based therapy in medical applications. Several researchers tried to improve the healing process by non-coherent light at different wavelengths. Table 5 lists the related works that deal with non-coherent light sources. Early studies by Pinheiro et al [93] irradiated four equidistant points with laser light (685 nm) or illuminated with wide range polarized light (400-2000 nm), both with doses of 20-40 J cm<sup>-2</sup>. Wounds treated by laser therapy with a dose of 20 J cm<sup>-2</sup> showed mild hyperemia, inflammation varied from moderate to intense, larger number of myofibroblasts without re-epithelialization. By increasing the dose to 40 J cm<sup>-2</sup>, exuberant neovascularization, severe hyperemia, moderate to severe inflammation, large collagen deposition and fewer myofibroblasts were observed. As a result, 685 nm laser therapy is capable of increasing collagen deposition and better organization on healing wounds. The number of myofibroblast was increased by using polarized light with low dosage exposure.

Blue LED has great potential as light therapy for wound healing. It can significantly influence biological systems, improving perfusion by releasing nitric oxide from nitrosyl complexes with hemoglobin in a skin flap model in rats. A comparison between red LED (630 nm) and blue LED (470 nm) has been reported by Adamskaya *et al* [43]. Although blue light does not penetrate tissue as deep as red light, blue light significantly contracts wound area and decreases keratin-1 mRNA based on planimeter measurement and histology analysis.

In more complicated matters, debate has occurred regarding determining whether the coherent and monochromatic laser is a better performer than non-coherent light such as LED or a filtered lamp. Comparisons have been made which prove that LED has yielded a more beneficial stimulation effect than LLLT [76, 94]. The results have shown that the effects of a serial LED probe such as green (570 nm) and red (660 nm) were found significantly higher than a low-level light probe. Infrared LED and LLL source provided a higher number of cells than the control cultures but no significant statistical difference. According to the amount of proliferation, the green probe yielded a significantly higher number of cells than red, infrared and low-level laser. Other related work is also reported by Demidova-Rice et al [94]. A comparison study has been conducted between four different wavelengths in the ranges of red and near-infrared light centered at 635, 670, 720, and 820 nm and a coherent beam of 633 nm. An 830 nm light source has revealed the most pronounced results in stimulating wound healing. However, no significant difference is observed between non-coherent 635 nm and coherent 633 nm in stimulation action in the rat model.

#### 4.5. Combination wavelength

Previous studies mostly performed with a single light source. However, there are some findings indicating the positive effect in healing processes by combining two or more energy sources. The combination of different energy sources can comprise laser, ultrasound, ultraviolet, electric current, magnetic field and microwave. Related works dealing with combination sources are summarized in table 6. Papageorgiou et al [95] have carried out a study on acne vulgarism treatment. A comparison treatment was conducted by using blue light (415 nm), mixed blue and red light (415-660nm), cool white light and 5% benzoyl peroxide cream. The results showed that the laser treatment with mixed blue and red light was effective in inflammatory lesions. Guffey et al [96] exploited blue laser 405 nm combined with infrared laser 880 nm on staphylococcus aureus and pseudomonas aeruginosa to depress and reduce the number of bacteria colonies. They claimed that such a combination has shown the most effective way to kill both bacteria.

Generally, blue light is commonly used for bactericide. Combination red and infrared light has been reported by Braverman et al [97], who dealt with 72 rabbits. Heliumneon laser radiation (He-Ne; 632.8 nm) and pulsed infrared laser radiation (IR; 904 nm) were combined to irradiate skin wounds. The tensile strength for the laser treated groups was more significant than the non-irradiated group; however, there were no significant differences in statistical data between the laser group for wound healing, collagen area and epidermal growth. Similar work was also reported by Lievens [85], who studied regeneration of the lymphatic system during the process of wound healing by combining cw He-Ne 632 nm and pulse infrared laser 904 nm. The frequency of the pulsed infrared laser is 1000 Hz with an energy density of  $2.1 \text{ J cm}^{-2}$ . The energy density of He–Ne laser is 1.2 J cm<sup>-2</sup> to treat 500 mice. The treatment is carried out twice daily. Laser treatment has enhanced the adhesion, oedema and the lymph vessels, thus accelerating the vein regeneration process of blood and lymph vessels during wound healing. Simunovic et al [122] studied this using a human specimen. They claimed that the wound healing, pain relief and functional recovery of patients was significantly improved for the group of patients treated by LLLT compared to untreated patients.

The combination of 685 and 830 nm reveals increased collagen production and organization [7]. Better repair of wounds was found by using energy density of 20 J cm<sup>-2</sup> as compared to that with 50 J cm<sup>-2</sup>. Combining the light exposure is more effective than using a single laser. In contrast, a similar experiment [80] obtained different results. Histological analysis was used to investigate the systemic action and repair process of wounds produced on the backs of rats and treated with red, infrared or both lasers applied directly or indirectly to the wounds. The combined application of red and infrared lasers resulted in the most evident systemic effect on the repair of skin wounds produced in rats.

Recently wounds have been treated by combining two light sources comprised of 685 and 830 nm [98]. Lack of beneficial effect is revealed after irradiation with high energy density of  $22 \text{ J cm}^{-2}$ .

Laser Phys. 24 (2014) 083001

Table 5. Wound healing that involves non-coherent light sources.

Author	Target	Wavelength (nm)	Energy density (J cm <sup>-2</sup> )	Power (mW)	Schedule	Outcome
Vinck [121]	Embryonic chicken fibroblast	570	0.1	10	3 d	Improved healing
Vinck [76]	Fibroblast (old chicken embryos)	Laser 830 LED 570 660 950	1 0.1 0.53 0.53	40 10 80 160	3 d	Improved healing (but no difference with 830 nm laser)
Demidova-Rice [94]	139 mice	Laser 632.8 LED 635 670 720 820	1	_	_	820 nm optimum wavelength and 635 nm second advanced, no difference between 632.8 nm and 633 nm
Pinheiro [93]	30 Wistar rat	400-2000	20	—	Every other days for 7 d	Improved healing (more benefit than 685 nm)
Al-Watban [57]	893 SD rat	Laser 532 633 810 980 10600 LED 510–872	4.71	_	Three times per week	633 nm improved healing
Adamskaya [43]	Rat	470 630 (LED)	$0.5 \mathrm{J} \mathrm{cm}^{-2}$	1000	For 5 d	Blue light significantly influences wound healing

Table 6. Wound healing involving combination wavelengths.

Author	Target	Waveleng	gth (nm)	Energy density (J cm <sup>-2</sup> )	Powe	er, mW	Schedule	Outcome	Remark
Papageorgiou	107 patients	$415 \pm 20$	660±20		4.23	2.67	Daily for	Optimum dose	Better than
Lievens [85]	500 mice	632	904	(1.2+2.1)	5	6800	Twice daily	Improved healing	Compared to control
Simunovic [122]	74 patients	632.8	904 (pulse)	—	—	—	—	Improved healing	Compared to
Braverman [97]	72 rabbits	632.8	904 (pulse)	(1.65+8.25)	—	—	Daily for 21 d	No effect	No significant difference with
Noudeh [98]	20 rats	670	810	(10+12)	500	250	_	No effect	Single laser Compared to
Rodrigo [80]	36 Wistar rats	685	830	20	30	50	Single	Worse	control Most evident systemic effect
Mendez [75]	60 rats	685	830	20	35	35		Optimum dose improved healing	on healing Combination better than
Nussbaum [99]	20 patients	820	30 super- luminous diode	50	15		Three times weekly	No effect Worse	Compared with US/UVC

Combinations were also organized between light source and other techniques. For example, two light source treatments were compared between ultrasound and ultraviolet-C (US/UVC) [99]. The light group combined a beam of 820 nm (energy density of 120 J cm<sup>-2</sup>) with non-coherent light of 4 J  $cm^{-2}$  operating in pulse mode with a repetition rate of 5000 pulses per second (pps). In this study, humans were used as targets. The patients were exposed three times weekly with the combination of light sources; and five times weekly for the combination of ultrasound (US) group (3MHz) with



Figure 3. Dependence of optimum dose on wavelength.

ultraviolet-C group (250 nm). The combination of US/UVC group has shown an advantage in comparison to the light sources group. This is possibly due to the tissues being absorbent without selection of the optimum characteristic [99].

Overall, the combination of irradiation was found to be more effective in comparison with single irradiation. The longer wavelength indicates a significant difference and enhances the effective healing.

#### 5. Summary and future trend

Optical technology is a promising technique to replace conventional wound treatment. Less cream or any medicine applied on the wound is normal practice in a conventional method. Lasers and non-coherent light sources like LED can surpass traditional medicine, resulting in less pain, faster and simple treatment. Laser wound treatment is a non-destructive technique, which is non-touch, and directly illuminated on to the wound. As a result, the wound area is sterilized, killing the bacteria and enhancing the collagen production (this is an important agent in the healing process). No need to clean, apply medicine or bandages. Such treatment is very promising, economical and fast. From the many light sources that have been discussed earlier, blue and near-infrared lasers have shown better performance in wound healing process. Therefore, further investigation using these two light sources is needed to as well as maybe the potential to combine both of them for better treatment in wound healing. In general, the relationship between the optimum doses with respect to wavelength is shown in figure 3. There is an optimum dose that is at 25 J cm<sup>-2</sup> corresponding to a wavelength in the red region. The blue and near-infrared have the smallest optimum dose of about 1 J cm<sup>-2</sup> for wound healing treatment.

### 6. Conclusion

In summary, low-level laser therapy at the appropriate dosimetric parameter can provide an acceleration effect in wound healing. The bio-stimulatory effects were dependent on the energy density or doses and laser wavelengths. Different light sources had different interactions with wounds. The visible laser region is capable of accelerating the wound repair via enhanced proliferation of cells and a reduction in inflammation. With in vivo sampling, the appropriate dose appears to be in the range 1-5 J cm<sup>-2</sup>, corresponding to a wavelength of 632.8 nm. It becomes more effective when administered on a daily basis. In vitro sampling, the suitable dose to stimulate human fibroblast is in the range  $0.5-5.0 \text{ J cm}^{-2}$ . For an infrared laser, the frequency of administering therapy should be less than visible laser therapy and the energy dose around  $0.5-10.0 \text{ J} \text{ cm}^{-2}$ . There is still a lack of attention given to energy doses higher than 10J cm<sup>-2</sup>. The infrared laser will achieve optimum treatment with high power dose and short exposure time. Apparently, a combination technique may join the effects of antibacterial and anti-inflammatory action in order to accelerate the healing process. Larger differences between two wavelengths perhaps increases the stimulatory effects. Polarized light would be more effective than an unpolarized light source. Finally, it is better to highlight that there is no significant difference between laser and LED effects on wound healing processes.

#### Acknowledgments

The authors would like to thank the Malaysian Department of Higher Education for financial support with this project through GUP grant 02H79. Thanks are also extended to MyBrain15 for the scholarship given to Lau Pik Suan for pursuing her PhD program in UTM. Thanks also to Dr Regunathan A/L Villanayer from Hospital Sultanah Bahiyah (HSB) for his informal discussion regarding laser applications in medicine.

#### Reference

- [1] Singer A J and Clark R 1999 N. Engl. J. Med. 341 738
- [2] Broughton G , Janis J E and Attinger C E 2006 *Plast. Reconstr. Surg.* 117 12S
- [3] Martin P 1997 Science 276 75
- [4] Aukhil I 2000 Periodontol. 22 44
- [5] Cukjati D, Reberšek S and Miklavčič D 2001 Med. Biol. Eng. Comput. 39 263
- [6] Komarcević A 2000 Med. Pregl. 53 363
- [7] Mester E, Szende B and Gärtner P 1968 Radiobiol., Radiother. 9 621
- [8] Kleinjung T 2011 Textbook of Tinnitus (Berlin: Springer) pp 749–52
- [9] Glinkowski W 2001 Lasers Musculoskeletal System (Berlin: Springer) pp 188–98
- [10] Hawkins D and Abrahamse H 2005 Photomed. Laser Ther.
   23 251
- [11] Mester A and Mester A F 1988 Laser Optoelectronics in Medicine (Berlin: Springer) pp 731–5
- [12] Mester E, Mester A F and Mester A 1985 Lasers Surg. Med. 5 31
- [13] Lilge L, Tierney K and Nussbaum E 2000 J. Clin Laser Med. Surg. 18 235
- [14] Rocha A Jr, Vieira B, De Andrade L and Aarestrup F 2009 Photomed. Laser Surg. 27 303
- [15] Oliveira R F D et al 2008 Photomed. Laser Surg. 26 6
- [16] Schindl A et al 1998 Diabetes Care 21 580

- [17] Enwemeka C S 1988 J. Orthop. Sports Phys. Ther. 9 333
- [18] Fillipin L I et al 2005 Lasers Surg. Med. 37 293
- [19] Reddy G K 2004 J. Clin. Laser Med. Sur. 22 141
- [20] Hawkins D, Houreld N and Abrahamse H 2005 Ann. NY Acad. Sci. 1056 486
- [21] Hamblin M R and Demidova T N 2006 Proc. SPIE 6140 614001
- [22] Huang Y-Y, Chen A C-H, Carroll J D and Hamblin M R 2009 Dose Response 7 358
- [23] Weiss R A, Weiss M A, Geronemus R G and McDaniel D H 2004 J. Drugs Dermatol. 3 605
- [24] Sher N A et al 1991 Arch. Ophthalmol. **109** 491
- [25] Gottsch J et al 1991 Ophthalmol. 98 146
- [26] Carson C A, Taylor H R and Laser M E 1995 Arch. Ophthalmol. 113 431
- [27] Taylor H R, Guest C S, Kelly P, Alpins N A and Laser E 1993 Arch. Ophthalmol. 111 1621
- [28] Seiler T and McDonnell P J 1995 Surv. Ophthalmol. 40 89
- [29] Roberts R E, Selby J E and Biberman L M 1976 Appl. Opt. 15 2085
- [30] de Sousa A P C et al 2010 Photomed. Laser Surg. 28 547
- [31] Kiecolt-Glaser J K, Marucha P T, Mercado A, Malarkey W and Glaser R 1995 Lancet 346 1194
- [32] Rappolee D A, Mark D, Banda M J and Werb Z 1988 Science 241 708
- [33] Hunt T, Knighton D, Thakral K, Goodson W 3rd and Andrews W 1984 *Surgery* **96** 48
- [34] Chen L, Tredget E E, Wu P Y and Wu Y 2008 *PloS One* 3 e1886
- [35] Germain L, Jean A, Auger F A and Garrel D R J. Surg. Res. 57 268
- [36] Montesano R and Orci L 1988 Proc. Natl Acad. Sci. USA 85 4894
- [37] MoulinV et al 1996 Burns 22 359
- [38] Yu W, Naim J O and Lanzafame R J 1997 Lasers Surg. Med. 20 56
- [39] Prabhu V et al 2012 J. Biophoton. **5** 168
- [40] Al-Watban F A and Zhang X Y 1996 *Laser Ther.* **8** 127
- [41] Al-Watban F A and Zhang Z 1994 Laser Ther. 6 119–24
- [42] Al-Watban F A and Zhang X 1997 J. Clin. Laser Med. Surg. 15 209
- [43] Adamskaya N et al 2011 Injury 42 917
- [44] Poon V K, Huang L and Burd A 2005 J. Photochem. Photobiol. B: Biol. 81 1
- [45] Edwards R and Harding K G 2004 Curr. Opin. Infect. Dis. 17 91
- [46] GuffeyJ S and Wilborn J 2006 Photomed. Laser Ther. 24 684
- [47] Kana J S and Hutschenreiter G 1981 Arch. Surg. 116 293
- [48] Surinchak J S, Alago M L, Bellamy R F, Stuck B E and Belkin M 1983 Lasers Surg. Med. 2 267
- [49] Lyons R F *et al* 1987 *Ann. Plast. Sur.* 18 47
  [50] Hunter J, Leonard L, Wilson R, Snider G and Dixon J 1984
- Lasers Surg. Med. 3 285
- [51] Fretz P B and Li Z 1992 *Can. Vet. J.* **33** 650
- [52] Atabey A, Karademir S, Atabey N and Barutcu A 1995 Eur. J. Plast. Surg. 18 99
- [53] Ghamsari S M et al 1997 Vet. Sur. 26 114
- [54] Ribeiro M S et al 2004 J. Clin. Laser Med. Surg. 22 59
- [55] Al-Watban F A 2009 *Photomed. Laser Surg.* 27 127
- [56] Al-Watban F A, Zhang X Y and Andres B L 2007 Photomed. Laser Surg. 25 72
- [57] Al-Watban F A, Zhang X Y, Andres B L and Al-Anize A 2009 Photomed. Laser Surg. 27 269
- [58] Al-Watban F A and Andres B L 2003 J. Clin. Laser Med. Surg. 21 249
- [59] Al-Watban F A and Delgado G D 2005 Photomed. Laser Ther.
   23 245
- [60] Al-Watban F A and Zhang X 2004 J. Clin. Laser Med. Surg. 22 15
- [61] Hawkins D H and Abrahamse H 2006 Lasers Surg. Med. 38 74

- [62] Houreld N and Abrahamse H 2007 Photomed. Laser Surg. 25 78
- [63] Evans D H and Abrahamse H 2008 Photodermatol., Photoimmunol. Photomed. 24 199
- [64] Houreld N and Abrahamse H 2010 *Diabetes Technol. Ther*. 12 971
- [65] Nussbaum E L et al 2009 Lasers Surg. Med. 41 372
- [66] Sekhejane P R, Houreld N N and Abrahamse H 2011 *Photomed. Laser Surg.* **29** 521
- [67] Walker M D, Rumpf S, Baxter G D, Hirst D G and Lowe A S 2000 Lasers Surg. Med. 26 41
- [68] Ribeiro M A G et al 2009 Photomed. Laser Surg. 27 49
- [69] Pugliese L S, Medrado A P, Reis S R A and Andrade Z A 2003 Pesqui. Odontol. Bras. 17 307
- [70] Medrado A R, Pugliese L S, Reis S R A and Andrade Z A 2003 Lasers Surg. Med. 32 239
- [71] Reis S R et al 2008 Photomed. Laser Surg. 26 307
- [72] Danno K, Mori N, Toda K I, Kobayashi T and Utani A 2001 Photodermatol., Photoimmunol. Photomed. 17 261
- [73] Stolik S, Delgado J, Perez A and Anasagasti L 2000*J. Photochem. Photobiol. B: Biol.* 57 90
- [74] Grossman N, Schneid N, Reuveni H, Halevy S and Lubart R 1998 Lasers Surg. Med. 22 212
- [75] Mendez T M, Pinheiro A L, Pacheco M T, Nascimento P M and Ramalho L M 2004 J. Clin. Laser Med. Surg. 22 19
- [76] Vinck E M, Cagnie B J, Cornelissen M J, Declercq H A and Cambier D C 2003 Lasers Med. Sci. 18 95
- [77] Tikiz C et al 2010 Turkiye Klinikleri J. Med. Sci. 30 135
- [78] Yamany A A and Sayed H M 2012 J. Adv. Res. 3 21
- [79] Demir H, Balay H and Kirnap M 2004 J. Rehabil. Res. Dev. 41 147
- [80] Rodrigo S M et al 2009 Photomed. Laser Surg. 27 929
- [81] Rezende S B, Ribeiro M S, Núñez S C, Garcia V G and Maldonado E P 2007 J. Photochem. Photobiol. B: Biol. 87 145
- [82] Lanzafame R J et al 2004 Photomed. Laser Ther. 22 483
- [83] Skopin M D and Molitor S C 2009 Photodermatol., Photoimmunol. Photomed. 25 75
- [84] Kawalec J S, Hetherington V J, Pfennigwerth T C, Dockery D S and Dolce M 2004 J. Foot Ankle Surg. 43 214
   [85] Lisure Med. Sci. Col.
- [85] Lievens P 1991 Lasers Med. Sci. 6 193
- [86] Ribeiro M S, de Fátima Teixeira da Silva D, Maldonado E P, de Rossi W and Zezell D M 2002 J. Clin. Laser Med. Surg. 20 37
- [87] Laubach H J, Tannous Z, Anderson R and Manstein D 2006 Lasers Surg. Med. 38 142
- [88] Yu S Y et al 2006 Photodermatol. Photoimmunol. Photomed. 22 78
- [89] Romanos G E, Pelekanos S and Strub J R 1995 Lasers Surg. Med. 16 368
- [90] de Freitas A C, Pinheiro A L B, Gerardt de Oliveira M and Pedreira Ramalho L M 2002 J. Clin. Laser Med. Surg. 20 221
- [91] Sommer A P, Pinheiro A L, Mester A R, Franke R-P and Whelan H T 2001 J. Clin. Laser Med. Surg. 19 29
- [92] Anneroth G, Hall G, Ryden H and Zetterqvist L 1988 Br. J. Oral Maxillofac. Surg. 26 12
- [93] Pinheiro A L B, Pozza D H, Oliveira M G D, Weissmann R and Ramalho L M P 2005 Photomed. Laser Ther. 23 485
- [94] Demidova-Rice T N, Salomatina E V, Yaroslavsky A N, Herman I M and Hamblin M R 2007 Lasers Surg. Med. 39 706
- [95] Papageorgiou P, Katsambas A and Chu A 2000 Br. J. Dermatol. 142 973
- [96] Guffey J S and Wilborn J 2006 Photomed. Laser Ther. 24 680
- [97] Braverman B et al 1989 Lasers Surg. Med. 9 50
- [98] Jahangiri Noudeh Y, Shabani M, Vatankhah N, Hashemian S J and Akbari K 2010 Photomed. Laser Surg. 28 621
- [99] Nussbaum E L, Biemann I and Mustard B 1994 Phys. Ther. 74 812

- [100] Núñez S C, Nogueira G E, Ribeiro M S, Garcez A S and Lage-Marques J L 2004 Lasers Surg. Med. 35 363
- [101] Rabelo S B et al 2006 Photomed. Laser Ther. 24 474
- [102] Yasukawa A, Hrui H, Koyama Y, Nagai M and Takakuda K 2007 J. Vet. Med. Sci./Japan Soc. Vet. Sci 69 799
- [103] Hegde V N *et al* 2011 *Photochem. Photobiol.* **87** 1433
- [104] Do Nascimento P M, Pinheiro A L B, Ângelo Castilho Salgado M and Pedreira Ramalho L M 2004 Photomed. Laser Ther. 22 513
- [105] Gal P et al 2006 Photomed. Laser Ther. 24 480
- [106] Medrado A P, Soares A P, Santos E T, Reis S R A and Andrade Z A 2008 J. Photochem. Photobiol. B: Biol. 92 144
- [107] de Oliveira Guirro E C, de Lima Montebelo M I, de Almeida Bortot B, da Costa Betito Torres M A and Polacow M L O 2010 Photomed. Laser Surg. 28 629
- [108] D'Arcangelo C et al 2007 Oral Surg., Oral Med., Oral Pathol., Oral Radiol., Endod. 103 764
- [109] Güngörmüş M and Akyol U 2009 Photomed. Laser Surg. 27 895
- [110] Souil E et al 2001 Br. J. Dermatol. 144 260
- [111] Petersen S, Botes C, Olivier A and Guthrie A 1999 Equine Vet. J. **31** 228

- [112] Lowe A S, Walker M D, O'Byrne M, Baxter G D and Hirst D G 1998 Lasers Surg. Med. 23 291
- [113] Ezzati A, Bayat M and Khoshvaghti A 2010 Photomed. Laser Surg. 28 603
- [114] Longo L, Evangelista S, Tinacci G and Sesti A G 1987 Lasers Surg. Med. 7 444
- [115] Skinner S, Gage J, Wilce P and Shaw R 1996 Aust. Dent. J. 41 188
- [116] Pereira A N, Eduardo C D P, Matson E and Marques M M 2002 Lasers Surg. Med. 31 263
- [117] Herascu N, Velciu B, Calin M, Savastru D and Talianu C 2005 Photomed. Laser Ther. 23 70
- [118] Silveira P C, Streck E L and Pinho R A 2007 J. Photochem. Photobiol. B: Biol. 86 279
- [119] Silveira P C *et al* 2009 *J. Photochem. Photobiol. B: Biol.* **95** 89
- [120] Sanati M H, Torkaman G, Hedayati M and Dizaji M M 2011 J. Photochem. Photobiol. B: Biol. 103 180
- [121] Vinck E M, Cagnie B J, Cornelissen M J, Declercq H A and Cambier D C 2005 *Photomed. Laser Ther.* **23** 167
- [122] Simunovic Z, Ivankovich A D and Depolo A 2000 J. Clin. Laser Med. Surg. 18 67